

REVIEW

MECHANISMS IN ENDOCRINOLOGY

Vitamin D and fertility: a systematic review

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Abstract

Background: Vitamin D has been well-known for its function in maintaining calcium and phosphorus homeostasis and promoting bone mineralization. There is some evidence that in addition to sex steroid hormones, the classic regulators of human reproduction, vitamin D also modulates reproductive processes in women and men.

Aim: The aim of this review was to assess the studies that evaluated the relationship between vitamin D and fertility in women and men as well as in animals.

Methods: We performed a systematic literature search in Pubmed for relevant English language publications published until October 2011.

Results and discussion: The vitamin D receptor (VDR) and vitamin D metabolizing enzymes are found in reproductive tissues of women and men. *Vdr* knockout mice have significant gonadal insufficiency, decreased sperm count and motility, and histological abnormalities of testis, ovary and uterus. Moreover, we present evidence that vitamin D is involved in female reproduction including IVF outcome (clinical pregnancy rates) and polycystic ovary syndrome (PCOS). In PCOS women, low 25-hydroxyvitamin D (25(OH)D) levels are associated with obesity, metabolic, and endocrine disturbances and vitamin D supplementation might improve menstrual frequency and metabolic disturbances in those women. Moreover, vitamin D might influence steroidogenesis of sex hormones (estradiol and progesterone) in healthy women and high 25(OH)D levels might be associated with endometriosis. In men, vitamin D is positively associated with semen quality and androgen status. Moreover, vitamin D treatment might increase testosterone levels. Testiculopathic men show low *CYP21R* expression, low 25(OH)D levels, and osteoporosis despite normal testosterone levels.

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Introduction

Vitamin D has been well-known for its function in maintaining calcium and phosphorus homeostasis and promoting bone mineralization (1). Mounting evidence suggests that hypovitaminosis D is linked to an increased risk for cancer (2), autoimmune diseases, diabetes, and cardiovascular diseases (1, 2, 3), indicating the importance of sufficient vitamin D levels. There is some evidence that in addition to sex steroid hormones, the classic regulators of reproduction, vitamin D also modulates reproductive processes in women and men.

Infertility is a complex disorder with significant medical, psychosocial, and economic aspects (4), which affects about 15% of couples (5). One major cause of female infertility is polycystic ovary syndrome (PCOS). Women affected by PCOS frequently suffer from oligo- or anovulation as well as obesity and insulin resistance. Population-based studies found that in 30–40% of infertile couples the underlying cause is the male factor (6). In this context it should be mentioned that the overall semen quality of men is

decreasing, which might partly be explained by environmental factors (7). Indeed, as much as 20% of young men have sperm concentration below the WHO recommendation level and 40% present with sperm concentrations below a level that is considered optimal for fertility (8). Moreover, many adverse aspects of male aging have been attributed to the decrease in testosterone levels (9) and some evidence suggests an association of androgen and vitamin D metabolism (10, 11).

The focus of this review is the examination of research evidence relating to vitamin D status and fertility as well as endocrine disturbances in women including IVF outcome, PCOS, and endometriosis as well as androgen levels and semen quality in men. Due to the relative paucity of data obtained from humans, this review includes discussion of animal data on the role of vitamin D in reproduction. The role of vitamin D in pregnancy, the perinatal period, and lactation has been comprehensively reviewed elsewhere (12, 13) and is not included in detail here.

We performed a systematic literature search in Pubmed for relevant English language publications

published until October 2011. We used the following search terms: 'vitamin D' and 'fertility', 'vitamin D' and 'reproduction', 'vitamin D' and 'PCOS'. In addition, we also used the search terms '25-hydroxyvitamin D (25(OH)D)', '1,25-dihydroxyvitamin D', and 'calcitriol' instead of vitamin D. We also used listed references from selected articles to expand the search.

Vitamin D metabolism

Vitamin D is a steroid hormone. The vitamin D precursor 7-dehydrocholesterol is a normal intermediary in the cholesterol pathway and is present in the skin (1). u.v.-B radiation induces the conversion of 7-dehydrocholesterol to provitamin D₃, which spontaneously isomerizes to cholecalciferol (vitamin D₃) (1). Vitamin D₃ is released into circulation and transported by the vitamin D-binding protein (VDBP). Approximately 80–90% derives from sunlight-induced production in the skin. A small amount of the body's total vitamin D is also derived from diet and/or supplements. This may derive from plants or fungi containing ergocalciferol (vitamin D₂) or fatty fish or cod-liver oil containing vitamin D₃ (1). Vitamin D from the skin and diet is metabolized in the liver by the enzyme 25-hydroxylase (encoded by *CYP2R1*) to 25(OH)D, which is used to determine a patient's vitamin D status into vitamin D sufficient (25(OH)D ≥ 30 ng/ml; multiply by 2.496 to convert nanograms per milliliter to nanomoles per liter), vitamin D insufficient (25(OH)D 20–29 ng/ml), and vitamin D deficient (25(OH)D < 20 ng/ml) (1). However, the 2011 report from the Institute of Medicine (IOM) recommends a 25(OH)D level of at least 20 ng/ml (50 nmol/l) based on positive vitamin D effects on bone health (14). 25(OH)D is metabolized in the kidneys by the enzyme 1 α -hydroxylase to its active form, 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃). The enzyme 1 α -hydroxylase is also found in many other tissues allowing the local conversion of 25(OH)D to the active 1,25(OH)₂D₃ (1).

Biological actions of vitamin D are mediated through the vitamin D receptor (VDR) that is distributed across various tissues including skeleton and parathyroid glands as well as reproductive tissues (1, 15) (see Fig. 1a for classical vitamin D effects). Vitamin D binds to the nuclear VDR, which then heterodimerizes with the retinoid X receptor. This in turn binds to the vitamin D responsive element located in the promoter regions of the target genes (16). The VDR interacts with other transcription factors such as coactivator proteins and transcription integrators such as calcium-binding proteins (17). This genomic pathway leading to changes in gene transcription takes hours to days (18). Another pathway is the interaction with a cell surface receptor and second messengers, leading to a more rapid response taking seconds to minutes (1, 18). Catabolization of 1,25(OH)₂D₃ and 25(OH)D to biologically inactive calcitric acid is catalyzed by 24-hydroxylase (1).

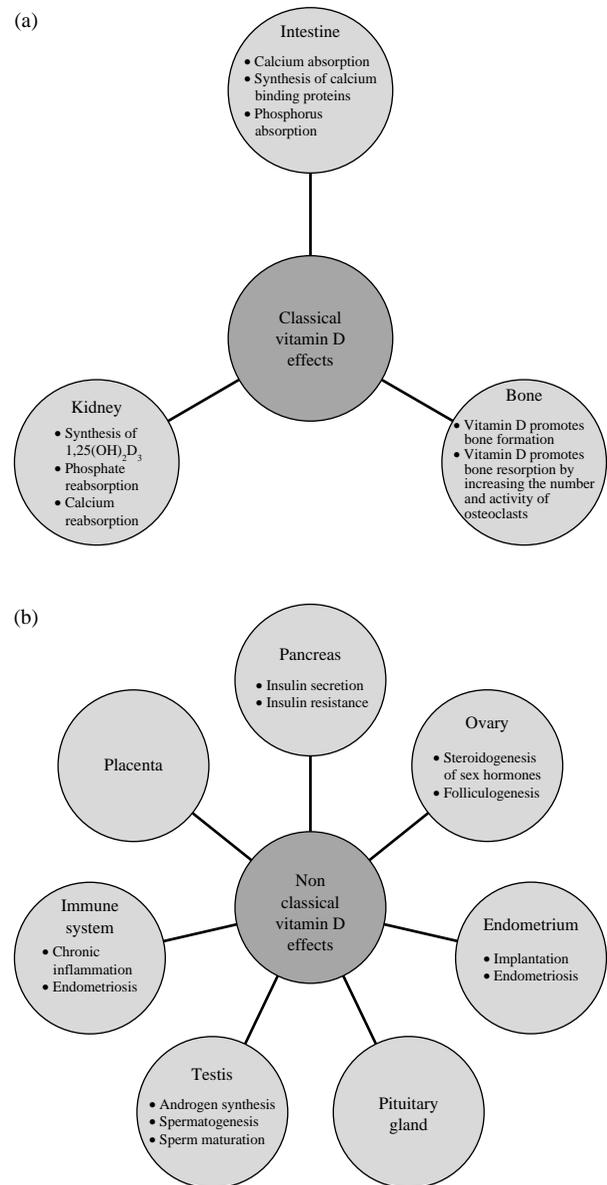


Figure 1 (a) Classical vitamin D effects. (b) Nonclassical vitamin D effects on various tissues (related to human fertility). 1,25(OH)₂D₃, 1,25-dihydroxyvitamin D₃.

VDR expression in reproductive tissues

Females

The VDR is distributed across various tissues suggesting an active role of vitamin D in those tissues (see also Fig. 1b for nonclassical vitamin D effects). In women, VDR mRNA has been shown to be expressed in the ovaries (19), in mixed ovarian cells, and in purified granulosa cell cultures indicating a role in steroidogenesis of sex hormones (20). Likewise, the human placenta expresses *CYP27B1* (encoding 1 α -hydroxylase) and

VDR (21, 22). Moreover, VDR has been found in the human pituitary gland (23) as well as in human endometrium (19). Viganò *et al.* (24) demonstrated by the measurement of $1,25(\text{OH})_2\text{D}_3$ levels in the supernatant of endometrial cells treated with vitamin D that endometrium is capable of extrarenal synthesis of the active form of vitamin D. The authors also showed that the active form of the 1α -hydroxylase gene was expressed in human endometrial stromal cells independent of the cycle phase but with a significant increase in early pregnant decidua (24).

Males

In the genital tract of male rodents, VDR has been found in the smooth muscles of the epididymis, spermatogonia, and Sertoli cells, indicating a role of vitamin D in spermatogenesis and sperm maturation in rats (25, 26). Moreover, the VDR was detected in human testicular tissue homogenates using titrated vitamin D (27). More recently, the VDR was detected in human sperm, with binding sites in the nucleus and the mid-piece of the sperm (28). An ultrastructural localization of VDR in human sperm showed that the VDR is largely located in the sperm nucleus (29). Recently, it was reported that VDR and vitamin D metabolizing enzymes are concomitantly expressed in spermatids, vesicles within the caput epididymis, and glandular epithelium of cauda epididymis, seminal vesicle, and prostate (30).

Vitamin D effects in reproductive tissues

Females

Vitamin D has a biologically plausible role in female reproduction. In human ovarian tissue, $1,25(\text{OH})_2\text{D}_3$ stimulated progesterone production by 13%, estradiol production by 9%, and estrone production by 21% (20). Likewise, it was demonstrated in a choriocarcinoma cell line that P450 aromatase activity (catalyzing the biosynthesis of estrogens) and expression are stimulated by calcitriol and that an atypical vitamin D response element is located in *CYP19* (*CYP19A1*) (encoding P450 aromatase) promoter (31). $1,25(\text{OH})_2\text{D}_3$ regulates human chorionic gonadotropin expression and secretion in human syncytiotrophoblasts (32) and increases placental sex steroid production (33). Previous studies have demonstrated that calcitriol promotes calcium transport in the placenta (34), stimulates placenta lactogen expression (35), and regulates *HOXA10* expression in human endometrial stroma cells (36). *HOXA10* expression is important for the development of the uterus and essential for endometrial development, allowing uterine receptivity to implantation (37).

Males

The specific mechanisms by which vitamin D influences male reproduction remain unclear. Recently, it has been shown that vitamin D treatment upregulates certain testis-specific genes in mouse testis (38), whereby 19 out of 2483 testis-specific genes showed upregulation by $1,25(\text{OH})_2\text{D}_3$ treatment. Of these genes, the regulator of cellular cholesterol homeostasis ATP-binding cassette transporter 1 (*ABCA1*) was expressed mainly in Sertoli cells and might influence male fertility. *Abca1* knockout mice have significantly reduced intratesticular testosterone levels as well as reduced sperm counts compared with wild-type animals (39). Absence of *Abca1* results in depletion of lipids, including HDL-C from Leydig cells, and HDL-C is the primary source of cholesterol for steroidogenic tissues. Thus, Leydig cell function is partially compromised in the absence of *Abca1* resulting in less testosterone production and fewer spermatozoa generation in *Abca1* knockout mice (39).

Moreover, Zanatta *et al.* (40) reported that $1,25(\text{OH})_2\text{D}_3$ triggers plasma membrane-initiated actions by modulating calcium uptake and by altering gamma-glutamyl transpeptidase (GGTP) activity in immature rat testis. GGTP is involved in the synthesis of specific proteins known to be secreted by Sertoli cells. For more details, the genomic and nongenomic vitamin D effects on rat testis have been reviewed elsewhere (41).

It has been shown that testosterone downregulates the VDR in testis cells (42). In cultured human osteoblasts, androgens increase 1α -hydroxylase, a key enzyme in vitamin D metabolism which converts $25(\text{OH})\text{D}$ to the biologically more active form $1,25(\text{OH})_2\text{D}$ (43).

In rats, vitamin D showed a potent stimulatory effect on amino acid accumulation in 11-day old rat testis that can be blocked by cycloheximide. The authors' conclusion from this study was that vitamin D plays an important role in the testis by genomic effects that can be triggered by protein kinase A as well as by rapid responses involving $\text{Ca}^{2+}/\text{K}^{+}$ channels on the plasma membrane (44). In addition, Akerstrom & Walters (45) showed an increased calcium uptake in TM4 Sertoli cells shared by a nuclear receptor activity mediated through $1,25(\text{OH})_2\text{D}_3$, indicating that vitamin D influences testis function via dual response pathways.

In a study investigating human sperms at the molecular level, $1,25(\text{OH})_2\text{D}_3$ has an effect on cholesterol efflux, protein phosphorylation, and increased sperm survival (29). Thus, vitamin D might play an important role in the extratesticular maturation of sperm by influencing capacitation and might modulate sperm survival. More recently, Aquila *et al.* (46) demonstrated that $1,25(\text{OH})_2\text{D}_3$ through VDR increased intracellular Ca^{2+} levels, motility, and acrosin activity revealing an effect of vitamin D in the

acquisition of fertilizing ability in human sperm. Moreover, $1,25(\text{OH})_2\text{D}_3$ reduces triglyceride content concomitantly to the increase of lipase activity (46) through the VDR. The authors speculate that lipid metabolism increases to meet the energetic demands during the process of capacitation by reducing energy storage and increasing energy expenditure.

Vitamin D and fertility in animal studies

In the following, we summarize evidence on the effect of vitamin D status on reproductive outcomes in animals.

With respect to the important role of vitamin D in calcium metabolism, it should be mentioned that animal investigations have established a role of calcium in oocyte activation and maturation, resulting in the resumption and progression of follicular development (47). Results from an *in vivo* study showed that $1,25(\text{OH})_2\text{D}_3$ significantly increases uterine weight and induces decidual reaction (48), suggesting a physiological role in endometrial cell differentiation into decidual cells, a crucial step in the process of blastocyst implantation. However, very high doses of $1,25(\text{OH})_2\text{D}_3$ result in a reduced corpus luteum, reduced progesterone, and alterations in the estrous cycle in rats (49). In diabetic rats, $1,25(\text{OH})_2\text{D}_3$ treatment has a protective effect on alloxan-induced damage in the reproductive system by enhancing testosterone and 17β -estradiol levels, consequently protecting from oxidative stress, cellular toxicity, and maintaining the number and motility of spermatozooids (50).

Vitamin D-deficient animals

Female rats It has been shown that vitamin D deficiency reduces mating success and fertility in female rats. In detail, female rats fed a vitamin D-deficient diet are capable of reproduction, but overall fertility is reduced including the probability of impregnation as well as an increased risk of pregnancy complications. This is not corrected by normalizing the hypocalcemia in vitamin D-deficient female rats, but requires $1,25(\text{OH})_2\text{D}_3$ (51).

Male rats In vitamin D-deficient male rats, it has been shown that although capable of reproducing, animals have a 45% reduction in successful matings as well as a decreased overall fertility rate that is reduced by 73% when compared with controls (52). The testes of vitamin D-deficient rats showed incomplete spermatogenesis and degenerative changes (53). Uhland *et al.* (54) demonstrated that the replacement of calcium alone in vitamin D-depleted animals was enough to restore fertility in male rats.

Vdr and 1α -hydroxylase knockout animals

The development of *Vdr* and 1α -hydroxylase knockout animals contributed much to the current knowledge about vitamin D and its role in reproductive function. With respect to steroidogenesis of sex hormones, in *Vdr* knockout mice, the aromatase activity in the ovary, testis, and epididymis is 24, 58, and 35% of the wild-type values respectively, and the gene expression of aromatase is also reduced. Moreover, elevated LH and FSH levels indicate hypergonadotropic hypogonadism. Supplementation of estradiol normalizes histological abnormality in the male and female gonads, whereas calcium supplementation increases aromatase activity and partially corrects hypogonadism (15). Interestingly, estrogen supplementation increased the uterine weight of female *Vdr* knockout mice indicating that vitamin D plays an important role in estrogen production in the ovary (55).

Both *Vdr* and 1α -hydroxylase knockout female mice conceive infrequently, have significantly fewer viable fetuses *in utero* that are also of lower body weight, and present with uterine hypoplasia, impaired folliculogenesis, anovulation, and absent corpora lutea (36, 55, 56, 57). In *Vdr* null mutant mice, feeding high calcium diets partly restores fertility (15, 56, 58) and increases the rate of conception but does not normalize the number or weight of viable fetuses. Thus, not only the lack of a direct effect of $1,25(\text{OH})_2\text{D}_3$ on reproductive function causes infertility, but also hypocalcemia has a role to play. Likewise, 1α -hydroxylase knockout female mice develop infertility accompanied by decreased estrogen and progesterone levels, elevated FSH and LH levels, defects in follicular development and corpus luteum formation as well as uterine hypoplasia (59). Again, the defective reproductive phenotype was normalized when the mice were fed a rescue diet (containing high calcium, phosphate, and lactose). The authors concluded that the infertility seen in $1,25(\text{OH})_2\text{D}_3$ -deficient mice is not a direct effect of active vitamin D deficiency on the reproductive system but rather an indirect effect mediated by extracellular calcium and phosphorus.

In male animals, *Vdr* null mutant mice present with significant gonadal insufficiency, with decreased sperm count and motility, and histological abnormalities of the testis (15). Interestingly, the reproductive organs of male 1α -hydroxylase null mutant mice appeared grossly normal (57).

Vitamin D and fertility in humans

Correlations between season and light with reproduction have been extensively investigated. Whereas nearly all focus on effects of visible light through retino-hypothalamic connections or through melatonin secreted during the absence of visible light, the role of

vitamin D in the explanation has been hardly recognized. Indeed, several studies suggest an association of vitamin D with fertility in humans.

Serum levels of 25(OH)D show a seasonal variation with high levels in summer and autumn and lower levels in winter and spring. In northern countries, where a strong seasonal contrast in luminosity exists, the conception rate is decreased during the dark winter months, whereas a peak in conception rate during summer leading to a maximum in birth rate in spring has been observed (60). Moreover, ovulation rates and endometrial receptivity seem to be reduced during long dark winters in northern countries (61). There are several possible explanations for these findings including altered hypothalamic–pituitary axis, brain neurotransmitters such as serotonin, dopamine, and endogenous opioids (60). Melatonin has long been investigated for this purpose with no direct effects. However, this fact might also be partly explained by the seasonal variation of vitamin D levels, which might influence several pathways including altered endometrial development and altered oocyte development.

Women

There is evidence that vitamin D exerts some effects on female reproduction including IVF outcome, PCOS, and endometriosis as well as on steroidogenesis in healthy women (see also Fig. 2). We discuss these vitamin D effects in the following section.

IVF Studies investigating the association of vitamin D status with IVF outcome revealed inconsistent results. In a study among 84 infertile women undergoing IVF, women with higher levels of 25(OH)D in serum and follicular fluid were significantly more likely to achieve clinical pregnancy following IVF, and high vitamin D levels were significantly associated with improved

parameters of controlled ovarian hyperstimulation (62). In contrast, Aleyasin *et al.* (63) found no significant associations of 25(OH)D levels in serum and follicular fluid with IVF outcomes in a study including 82 infertile women undergoing assisted reproductive technology. Anifandis *et al.* (64) investigated 101 consecutive women who underwent 101 IVF–intracytoplasmic sperm injection (ICSI) ovarian stimulation cycles. In this study, women with a sufficient vitamin D status (25(OH)D > 30 ng/ml in follicular fluid) had a lower quality of embryos and were less likely to achieve clinical pregnancy when compared with women with insufficient (follicular fluid 25(OH)D 20.1–30 ng/ml) or deficient vitamin D status (follicular fluid 25(OH)D < 20 ng/ml). Altogether, to date, there is insufficient data to accurately evaluate the effects of vitamin D in women undergoing IVF.

Polycystic ovary syndrome PCOS is the most common female endocrine disorder with a prevalence of ~5–10% in women of reproductive age (65, 66, 67). PCOS is characterized by increased ovarian and adrenal androgen secretion, hyperandrogenic symptoms such as hirsutism, acne and/or alopecia, menstrual irregularity, and polycystic ovaries. In addition, insulin resistance is common in PCOS women (68), who are therefore at an increased risk of type 2 diabetes (69). Overall, PCOS is the most common cause of anovulatory infertility in women.

There is some evidence suggesting that vitamin D deficiency might be involved in the pathogenesis of insulin resistance and the metabolic syndrome in PCOS (70, 71), whether vitamin D is also related to endocrine parameters and fertility in PCOS is less clear. There are, however, several studies (70, 72, 73) correlating low 25(OH)D status with features of PCOS. In detail, in a study among 100 women with PCOS from Turkey, the authors observed a correlation of 25(OH)D levels with testosterone and DHEAS levels and the LH/FSH ratio (72). In contrast, in a study among 120 PCOS patients, Hahn *et al.* (73) found an association of 25(OH)D levels with free androgen index and SHBG but not with total testosterone, androstenedione, DHEAS, estradiol, or LH/FSH ratio. Of note, no adjustments for obesity or BMI were performed in that study (73). Another study among 206 women with PCOS revealed similar results showing an association of vitamin D with SHBG and hirsutism score but not with testosterone and free testosterone (70); the association of vitamin D with hirsutism score remained significant after adjusting for BMI but was attenuated when analyzing SHBG. Moreover, vitamin D deficiency was (BMI) independently associated with low insulin sensitivity assessed by quantitative insulin sensitivity check index in a study including 27 PCOS and 20 control women (71). In addition, vitamin D deficiency was found to be more common in PCOS women than in controls in an Iranian cohort including 85 PCOS and 115 control women (74),

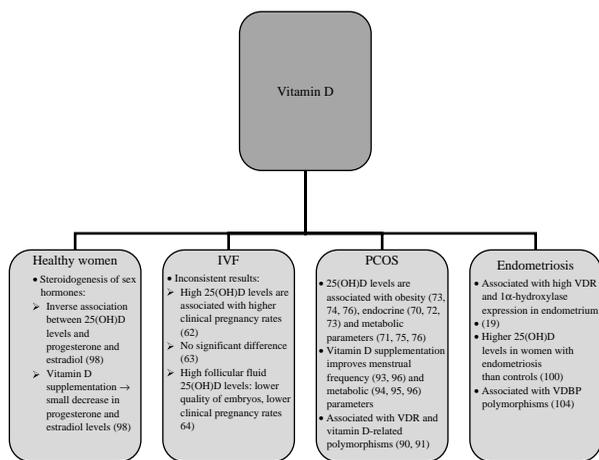


Figure 2 The proposed associations of vitamin D status with female reproduction.

as well as in a smaller observational study including 52 women (25 PCOS women and 27 controls) from Edinburgh (75). In the latter study, controls had a significantly lower BMI than PCOS women, which might explain the differences in 25(OH)D levels between PCOS and control women. However, the authors found that vitamin D deficiency was independently associated with lower insulin sensitivity and lower HDL-C levels (independent of BMI and waist-to-hip ratio) (75). Moreover, Panidis *et al.* (76) found an inverse association of 25(OH)D with obesity as well as a BMI-dependent association with insulin resistance in a cohort of 291 PCOS and 109 control women. In contrast, vitamin D intake was not associated with reported anovulatory infertility in the Nurses' Health Study II, a study prospectively following 18 555 married, premenopausal women without a history of infertility who attempted a pregnancy or became pregnant during an 8-year period, but there was no data on actual 25(OH)D levels (77).

The mechanisms underlying the association of low 25(OH)D levels and insulin resistance are not fully understood. As obesity is related to insulin resistance in PCOS (68) as well as in healthy subjects, the association of obesity with vitamin D deficiency deserves further discussion. So far, it is not clear whether vitamin D insufficiency results from obesity and/or if obesity is a consequence of vitamin D insufficiency. On the one hand, obesity may contribute to low circulating vitamin D levels by trapping vitamin D in fat tissues. Wortsman *et al.* (78) demonstrated that the increase of 25(OH)D levels 24 h after whole-body u.v.-light exposure was 57% lower in obese than in nonobese subjects. On the other hand, obese patients may avoid sunlight, which is necessary for the synthesis of vitamin D in the skin (79). This might be especially the case in hirsute PCOS women, who might tend to hide from the public due to their appearance.

There is evidence that low vitamin D levels are associated with obesity (73) and vice versa low vitamin D intake might be an independent predictor of obesity (80). The association of low vitamin D levels with insulin resistance might at least in part be mediated by obesity. Of note, some studies reported an association of low vitamin D status with insulin resistance only in obese PCOS women (72, 73, 76). Moreover, 25(OH)D levels have been reported to be lower in obese PCOS women when compared with normal weight PCOS women (73, 74, 76). Thus, proper studies should account for the role of obesity when investigating the association of vitamin D with insulin resistance.

There are, however, mechanisms beyond obesity that might explain the association of vitamin D deficiency with insulin resistance. First, vitamin D may have a beneficial effect on insulin action by stimulating the expression of insulin receptors and thereby enhancing insulin responsiveness for glucose transport (81). The vitamin D responsive element is present in the promoter

of the human insulin gene (82) and the transcription of the human insulin gene is activated by $1,25(\text{OH})_2\text{D}_3$ (83). Second, vitamin D regulates extracellular and intracellular calcium which is essential for insulin-mediated intracellular processes in insulin-responsive tissues such as skeletal muscle and adipose tissue (81). Moreover, alterations in calcium flux can have adverse effects on insulin secretion, which is a calcium-dependent process (84). Finally, as vitamin D has a modulating effect on the immune system (85), hypovitaminosis D might induce a higher inflammatory response, which is again associated with insulin resistance (86).

The use of statins in PCOS might improve not only lipids but also hyperandrogenemia (87). Interestingly, it has been shown in a randomized controlled trial (RCT) in 40 PCOS patients that 12-week treatment with atorvastatin at a dose of 20 mg daily resulted in a significant increase in serum 25(OH)D concentrations that was independent of the lipid-lowering effect of atorvastatin (88). Moreover, there was a significant correlation between the increases in 25(OH)D concentrations and the reduction of high-sensitive C-reactive protein.

The notion that vitamin D is involved in metabolic and endocrine parameters in PCOS is supported by the fact that the VDR regulates more than 3% of the human genome, including genes that are crucial for glucose metabolism (1, 81). In this context it has been shown that VDR-related polymorphisms (*Cdx2*, *Bsm-I*, *Fok-I*, *Apa-I*, and *Taq-I*) are related to vitamin D metabolism and might contribute to PCOS susceptibility (89). In detail, Mahmoudi (89) found an association of VDR *Apa-I* polymorphism with PCOS susceptibility in a cohort including 162 PCOS and 162 control women from Tehran.

In an Austrian cohort including 545 PCOS and 145 control women, the authors found an association of VDR *Cdx2* with insulin metabolism whereas the VDR *Apa-I* variant was associated with hyperandrogenemia (90). No association was found between VDR-related polymorphisms and PCOS susceptibility. Moreover, a smaller study including 56 PCOS women from Iran found an association of VDR *Taq-I* with elevated serum levels of LH as well as associations between decreased levels of SHBG and VDR *Bsm-I* (91). Recently, results from a genome-wide association study of 25(OH)D levels in 33 996 individuals of European descent from 15 cohorts established a role for common genetic variants in the regulation of 25(OH)D levels (92). Variants near genes involved in cholesterol synthesis (*DHCR7*), hydroxylation (*CYP2R1*), and vitamin D transport (*GC*) have been shown to affect vitamin D status, which has also been shown in PCOS women (90).

The evidence on the effects of vitamin D supplementation in PCOS women is sparse. There are, however, some small intervention trials showing

promising results. In a small-scale intervention study including 13 premenopausal women with chronic anovulation and hyperandrogenism, vitamin D repletion with ergocalciferol 50 000 U weekly or biweekly combined with administration of 1500 mg calcium daily resulted in normalization of menstrual cycles in seven women and two became pregnant (93). In addition, clinical improvements of acne vulgaris were observed in all three women affected by this condition (93). Of note, it is not possible to distinguish between calcium and vitamin D effects in that study. In contrast, in a pilot study among 13 obese women with PCOS, the administration of the single dose of 300 000 U vitamin D₃ orally did not significantly change BMI or the levels of DHEAS, total testosterone, free testosterone, and androstenedione levels but had a beneficial impact on insulin resistance assessed by homeostasis model assessment index (94). Similarly, in a small study including 15 obese PCOS women treated with 1 µg αcalcidol daily over 3 months, vitamin D treatment improved insulin secretion and had a beneficial effect on lipid status whereas BMI did not significantly change (95). In another study including 57 women who received 50 000 IU vitamin D₃ weekly over 24 weeks, vitamin D supplementation resulted in improved glucose metabolism as well in an improvement of menstrual frequency without a significant change of BMI (96). Likewise, in a study among 60 infertile PCOS women, metformin treatment combined with calcium and vitamin D supplementation resulted in a higher number of dominant follicles when compared with metformin alone and placebo, which might indicate a beneficial effect on fertility (97). Although these are very promising data derived from uncontrolled intervention studies, to date, there is no RCT to evaluate the effects of vitamin D treatment on endocrine and metabolic parameters in PCOS women. Considering the association of vitamin D deficiency with insulin resistance and type 2 diabetes in PCOS as well as in other cohorts (1, 81), and the association of poor vitamin D status with cardiovascular disease and fatal events (3), further research including large RCTs are highly warranted in this high-risk cohort. Moreover, the association of 25(OH)D levels with metabolic and endocrine parameters in PCOS women as well as the promising results from intervention studies in PCOS women might lead to a recommendation for measuring 25(OH)D and for vitamin D supplementation to improve fertility as well as metabolic disturbances.

Healthy women Evidence on the effects of vitamin D on fertility in healthy women is sparse. A study among 101 young volunteer women showed an inverse association between 25(OH)D levels and progesterone as well as estradiol (98). Likewise, in a study among 37 young volunteer women, four weekly doses of 28 000 IU vitamin D₃ resulted in a small decrease in estradiol and progesterone levels, although neither

change reached significance (98). As high 25(OH)D levels are associated with a reduced risk for breast cancer (99), the potential lowering effect of vitamin D on estradiol and progesterone levels might partly explain this association and deserves further investigation.

Endometriosis The pathogenesis of endometriosis is related to an impairment of immunologic mechanisms and inflammatory responses. Evidence on the association of endometriosis with vitamin D metabolism is sparse, but there are two points in favor for an association: i) it has been shown that the VDR and 1α-hydroxylase are expressed in the endometrium (24), suggesting that endometrium is an extrarenal site of vitamin D synthesis and vitamin D action; and ii) vitamin D is involved in the regulation of the immune system (1). Given that vitamin D is an effective regulator of the immune system and endometriosis has been shown to be associated with significant immune derangements, one might speculate about an influence of vitamin D in the local immune suppression and development of endometriosis. Of note, Agic *et al.* (19) found a significantly higher VDR and 1α-hydroxylase expression in the endometrium of women with endometriosis compared with healthy controls, which was also demonstrated at the protein level. However, 25(OH)D levels were similar in cases and controls. In a study among 87 women with endometriosis and 53 controls, Somigliana *et al.* (100) observed an association of high 25(OH)D levels with a significantly increased risk for endometriosis as well as a biological gradient showing higher levels of 25(OH)D in women with advanced stages of the disease. In contrast, in a study among 42 women with endometriosis and 113 control women, Hartwell *et al.* (101) found significantly higher 1,25(OH)D levels in women with endometriosis, whereas 25(OH)D levels were similar in both groups. More recently, Vilarino *et al.* (102) investigated the association of VDR polymorphisms with endometriosis and infertility in a cross-sectional study including 132 women with endometriosis-related infertility, 62 women with idiopathic infertility, and 133 controls. The authors found no association of VDR polymorphisms with endometriosis or infertility. In addition to 25(OH)D levels, VDBP was also found to be associated with endometriosis. Faserl *et al.* (103) found that the abundance of VDBP was higher in all endometriosis pools by a factor of ~3 compared with the control pool. Moreover, a polymorphism in the VDBP (GC-2) might be involved in the pathogenesis of endometriosis. Borkowski *et al.* (104) compared total concentrations of VDBP in serum and peritoneal fluid of women with and without endometriosis and found no differences. Another study investigating VDBP reported that one isoform of VDBP was significantly lower in the peritoneal fluid, but not in plasma, of women with endometriosis compared with controls (105).

Men

In men, vitamin D status might be related to spermatogenesis, semen quality, and testiculopathies as well as male hypogonadism (see also Fig. 3).

Semen and testiculopathy There is ample evidence showing that calcium is important in the male reproductive tract, where it is essential for spermatogenesis, sperm motility, hyperactivation, and acrosome reaction (106). However, the role of vitamin D, which is known as an important regulator of calcium metabolism, in semen quality and spermatogenesis is less clear and was the focus of several studies conducted in recent years. In detail, Blomberg Jensen *et al.* (107) studied the association of semen quality and vitamin D status in a cross-sectional study including 300 men from the general population. The authors found a positive correlation of 25(OH)D serum levels with sperm motility and progressive motility. Moreover, men with vitamin D deficiency (<10 ng/ml) had a lower proportion of motile, progressive motile, and morphologically normal spermatozoa compared with men with sufficient vitamin D status (>30 ng/ml). In addition to the cross-sectional analysis, Blomberg Jensen *et al.* (107) investigated the effect of 1,25(OH)₂D₃ on human sperm *in vitro* and found that 1,25(OH)₂D₃ increased intracellular calcium concentration in human spermatozoa through VDR-mediated calcium release from an intracellular calcium storage, increased sperm motility, and induced the acrosome reaction *in vitro*. In contrast, another study investigating the association of vitamin D status with semen quality in 307 young healthy men found a trend toward an association of high vitamin D levels with lower total sperm count and percentage of normal morphology sperm (108). However, those trends totally disappeared in the multivariate model adjusting for season, history of diseases of the reproductive organs, smoking of the young men, maternal smoking during pregnancy, maternal alcohol during pregnancy, abstinence time, and spillage during collection of the sample.

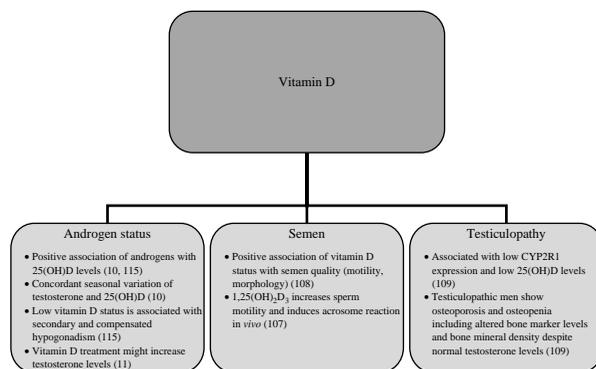


Figure 3 The proposed associations of vitamin D deficiency with male reproduction.

Moreover, Foresta *et al.* (109) examined CYP2R1 (encoding 25-hydroxylase) expression in 57 patients with testiculopathy (Sertoli cell only syndrome or severe hypospermatogenesis) and 41 controls. The authors found a significantly lower CYP2R1 gene and protein expression as well as significantly lower serum 25(OH)D levels in men with testiculopathy when compared with control subjects. Interestingly, testiculopathic patients showed osteopenia and osteoporosis including increased-bone marker levels and altered bone mineral density despite normal testosterone levels compared with controls. While there is some evidence from observational studies showing an association of vitamin D with semen quality, to date, there is no data from RCTs investigating the effects of vitamin D treatment on semen quality. However, in a phase II, randomized, double-blind, placebo-controlled trial, elocalcitol, a synthetic derivate of vitamin D₃, was administered for 3 months to 121 male patients with chronic prostatitis/chronic pelvic pain syndrome. Treatment with elocalcitol significantly reduced levels of IL8 in semen, suggesting an improved semen quality and forward motility of sperm (110). Unfortunately, further developments of elocalcitol, primarily designed for treatment of benign prostatic hyperplasia and overactive bladder, were terminated due to disappointing results regarding irritative urinary symptoms.

However, the discovery that 1,25(OH)₂D₃ influences sperm function may be useful for the development of novel therapeutic approaches to the treatment of male reproductive disorders.

Testosterone Low levels of vitamin D and androgens are both associated with increased mortality in men (3, 111, 112). Considering the fact that both low vitamin D and low testosterone levels are associated with obesity (1, 113), obesity might be a confounding factor when analyzing the association of low vitamin D or androgen status with mortality. However, in the above-mentioned studies (3, 111, 112), the association of vitamin D and androgens with increased mortality remained significant after adjustment for BMI. Interestingly, accumulating evidence exists for a complex interplay of vitamin D and androgen metabolism. It has been shown that androgens increase 1 α -hydroxylase (43). Furthermore, it was demonstrated that the regulation of gene expression by vitamin D metabolites is modified according to androgen levels (114). These data may suggest that androgen deficiency could hypothetically amplify the adverse health consequences of vitamin D deficiency.

Among 2299 men referred for coronary angiography, androgen levels and 25(OH)D level were independently associated and revealed a concordant seasonal variation (10). Likewise, results from the European Male Aging Study (EMAS) suggest an independent association of vitamin D status with compensated as well as secondary hypogonadism (115). Moreover, previous data indicate

that vitamin D therapy might increase testosterone levels (11) by yet largely unclear mechanisms. In detail, men undergoing a weight reduction program received either 83 µg (3332 IU) vitamin D daily for 1 year ($n=31$) or placebo ($n=23$). Compared with baseline values, a significant increase in total testosterone levels, bioactive testosterone, and free testosterone levels was observed in the vitamin D supplemented group. By contrast, there was no significant change in androgen levels in the placebo group. In view of the clinical significance of low testosterone and 25(OH)D levels we want to stress that further studies are needed to investigate the impact of vitamin D supplementation on androgen status in men and to evaluate the effect of testosterone replacement in men with respect to vitamin D status.

Pregnancy

The role of vitamin D is reviewed in detail elsewhere (13). In brief, it is well-known that vitamin D deficiency is prevalent among pregnant women (116) and pregnant women have significantly lower levels of 25(OH)D than nonpregnant control women (117). Approximately two in three pregnant women in the United States have suboptimal vitamin D status, with an even higher prevalence among black and Mexican-American women (118). Reduction of plasma 25(OH)D could contribute to the fall in plasma calcium during pregnancy, and may result from enhanced maternal metabolism or increased utilization of vitamin D by the fetus (117). Moreover, maternal vitamin D deficiency might be independently associated with an elevated risk for gestational diabetes mellitus (GDM) (119) and serum concentrations of 25(OH)D levels have been shown to be significantly lower in women with GDM than in pregnant women without GDM (120). In addition, vitamin D deficiency among pregnant women has been associated with elevated risk for other pregnancy complications such as preeclampsia (121) and bacterial vaginosis (122). In the Norwegian Mother and Child Cohort Study investigating 23 423 nulliparous pregnant women, low vitamin D intake was associated with an increased risk for the development of preeclampsia (123). Maternal serum 25(OH)D concentrations are associated with small-for-gestational age births in women (124, 125) as well as with offspring rickets (126), reduced bone density (127), asthma, (128) and schizophrenia (129). Recently, Hollis *et al.* (130) assessed the safety and effectiveness of vitamin D supplementation during pregnancy. In an RCT, women with a singleton pregnancy at 12–16 weeks' gestation received 400, 2000, or 4000 IU of vitamin D₃ per day until delivery. Not a single adverse event was attributed to vitamin D supplementation or circulating 25(OH)D levels. The authors concluded that vitamin D supplementation of 4000 IU/day for pregnant women is safe and most effective in achieving sufficiency in all women

and their neonates. Moreover, vitamin D supplementation resulted in a 50% reduction in preterm delivery, 25% reduction in infections of the mother, and 30% reduction in comorbidities (DM, hypertension, and preeclampsia) (131).

Vitamin D supplementation

To date, there are no specific guidelines regarding vitamin D supplementation for women or men affected by endocrine disturbances including infertility or hypogonadism. Thus, we will briefly discuss recent recommendations by the IOM (14) and the Endocrine Society (132).

The 2011 report from the IOM recommends a 25(OH)D level of at least 50 nmol/l (20 ng/ml) based on positive vitamin D effects on bone health (14). The report asserts that the daily vitamin D intake should be 600 IU for individuals up to the age of 70 years and 800 IU for older adults of the general population corresponding to the Recommended Daily Allowance (RDA, covering requirements of $\geq 97.5\%$ of the population). The tolerable upper intake level (UL) is defined as 4000 IU/day. The Endocrine Practice Guidelines Committee (132) suggests a daily intake of 1500–2000 IU vitamin D₃ daily for adults >18 years up to 70 years in order to raise the blood level of 25(OH)D consistently above 30 ng/ml; the UL was defined as 10 000 IU/day. The same daily intake and UL were suggested for adult pregnant and lactating women.

In general, vitamin D supplementation with 1000 IU/day increases 25(OH)D levels per 10 ng/ml (133). However, some authors recommend higher loading doses (e.g. 50 000 IU weekly for up to 8 weeks) in severely vitamin D-deficient subjects and those have been shown to be safe. Importantly, vitamin D intoxication resulting in hypercalcemia, renal damage, and vascular calcification is not observed until 25(OH)D levels >150 ng/ml (1).

Considering the multiple adverse effects of vitamin D deficiency on various health aspects, vitamin D supplementation in order to achieve a sufficient vitamin D status is of high importance.

Clinical consequences and summary

We presented evidence that vitamin D deficiency might be important for endocrine disturbances including fertility in women as well as in men. To date, the evidence is based largely on animal work and observational studies rather than on intervention trials. Nevertheless, there are some promising findings deserving further investigation.

Given the high prevalence of infertility as well as vitamin D insufficiency in otherwise healthy young women (70) and men (134) and the possible role of

vitamin D in human reproduction, research might lead to new therapeutic approaches such as vitamin D supplementation in the treatment of female and male reproductive disorders. Recognizing the fact that sperm quality is decreasing and infertility is a problem affecting about 10–15% of couples, this review might provide a rationale for further research in this area. We want to emphasize the fact that in infertility cases drastic improvements in reproductive failure may not be achieved by vitamin D treatment alone; however, vitamin D supplementation is a safe and cheap treatment, which might have some beneficial effects on human reproduction. Moreover, the impact of vitamin D supplementation on androgen levels deserves further investigation with respect to the adverse consequences of both vitamin D deficiency and male hypogonadism.

High-quality RCTs with a large sample size are required to determine the optimal 25(OH)D levels and to evaluate the effect of vitamin D supplementation on fertility.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References

- Holick MF. Vitamin D deficiency. *New England Journal of Medicine* 2007 **357** 266–281. (doi:10.1056/NEJMr070553)
- Pilz S, Dobnig H, Winkhofer-Roob B, Riedmüller G, Fischer JE, Seelhorst U, Wellnitz B, Boehm BO & März W. Low serum levels of 25-hydroxyvitamin D predict fatal cancer in patients referred to coronary angiography. *Cancer Epidemiology, Biomarkers and Prevention* 2008 **17** 1228–1233. (doi:10.1158/1055-9965.EPI-08-0002)
- Pilz S, März W, Wellnitz B, Seelhorst U, Fahrleitner-Pammer A, Dimai HP, Boehm BO & Dobnig H. Association of vitamin D deficiency with heart failure and sudden cardiac death in a large cross-sectional study of patients referred for coronary angiography. *Journal of Clinical Endocrinology and Metabolism* 2008 **93** 3927–3935. (doi:10.1210/jc.2008-0784)
- Benyamini Y, Gozlan M & Kokia E. Variability in the difficulties experienced by women undergoing infertility treatments. *Fertility and Sterility* 2005 **83** 275–283. (doi:10.1016/j.fertnstert.2004.10.014)
- Thonneau P, Marchand S, Tallec A, Ferial ML, Ducot B, Lansac J, Lopes P, Tabaste JM & Spira A. Incidence and main causes of infertility in a resident population (1,850,000) of three French regions (1988–1989). *Human Reproduction* 1991 **6** 811–816.
- Forti G & Krausz C. Clinical review 100: evaluation and treatment of the infertile couple. *Journal of Clinical Endocrinology and Metabolism* 1998 **83** 4177–4188. (doi:10.1210/jc.83.12.4177)
- Gyllenborg J, Skakkebaek NE, Nielsen NC, Keiding N & Giwercman A. Secular and seasonal changes in semen quality among young Danish men: a statistical analysis of semen samples from 1927 donor candidates during 1977–1995. *International Journal of Andrology* 1999 **22** 28–36. (doi:10.1046/j.1365-2605.1999.00137.x)
- Jørgensen N, Asklund C, Carlsen E & Skakkebaek NE. Coordinated European investigations of semen quality: results from studies of Scandinavian young men is a matter of concern. *International Journal of Andrology* 2006 **29** 54–61. (doi:10.1111/j.1365-2605.2005.00635.x)
- Zitzmann M. Testosterone deficiency, insulin resistance and the metabolic syndrome. *Nature Reviews. Endocrinology* 2009 **5** 673–681. (doi:10.1038/nrendo.2009.212)
- Wehr E, Pilz S, Boehm BO, März W & Obermayer-Pietsch B. Association of vitamin D status with serum androgen levels in men. *Clinical Endocrinology* 2010 **73** 243–248. (doi:10.1111/j.1365-2265.2010.03852.x)
- Pilz S, Frisch S, Koertke H, Kuhn J, Dreier J, Obermayer-Pietsch B, Wehr E & Zittermann A. Effect of vitamin D supplementation on testosterone levels in men. *Hormone and Metabolic Research* 2011 **43** 223–225. (doi:10.1055/s-0030-1269854)
- Kovacs CS. Vitamin D in pregnancy and lactation: maternal, fetal, and neonatal outcomes from human and animal studies. *American Journal of Clinical Nutrition* 2008 **88** 520S–528S.
- Lewis S, Lucas RM, Halliday J & Ponsonby AL. Vitamin D deficiency and pregnancy: from preconception to birth. *Molecular Nutrition and Food Research* 2010 **54** 1092–1102.
- Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA, Gallagher JC, Gallo RL, Jones G, Kovacs CS, Mayne ST, Rosen CJ & Shapses SA. The 2011 Report on Dietary Reference Intakes for Calcium and Vitamin D from the Institute of Medicine: What Clinicians Need to Know. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** 53–58. (doi:10.1210/jc.2010-2704)
- Kinuta K, Tanaka H, Moriwake T, Aya K, Kato S & Seino Y. Vitamin D is an important factor in estrogen biosynthesis of both female and male gonads. *Endocrinology* 2000 **141** 1317–1324. (doi:10.1210/en.141.4.1317)
- Jones G, Strugnell SA & DeLuca HF. Current understanding of the molecular actions of vitamin D. *Physiological Reviews* 1998 **78** 1193–1231.
- Jenster G, Spencer TE, Burcin MM, Tsai SY, Tsai MJ & O'Malley BW. Steroid receptor induction of gene transcription: a two-step model. *PNAS* 1997 **94** 7879–7884. (doi:10.1073/pnas.94.15.7879)
- Bouillon R, Carmeliet G, Verlinden L, van Etten E, Verstuyf A, Luderer HF, Lieben L, Mathieu C & Demay M. Vitamin D and human health: lessons from vitamin D receptor null mice. *Endocrine Reviews* 2008 **29** 726–776. (doi:10.1210/er.2008-0004)
- Agic A, Xu H, Altgassen C, Noack F, Wolfler MM, Diedrich K, Friedrich M, Taylor RN & Hornung D. Relative expression of 1,25-dihydroxyvitamin D₃ receptor, vitamin D 1 α -hydroxylase, vitamin D 24-hydroxylase, and vitamin D 25-hydroxylase in endometriosis and gynecologic cancers. *Reproductive Sciences* 2007 **14** 486–497. (doi:10.1177/1933719107304565)
- Parikh G, Varadinova M, Suwandhi P, Araki T, Rosenwaks Z, Poretsky L & Seto-Young D. Vitamin D regulates steroidogenesis and insulin-like growth factor binding protein-1 (IGFBP-1) production in human ovarian cells. *Hormone and Metabolic Research* 2010 **42** 754–757. (doi:10.1055/s-0030-1262837)
- Tanamura A, Nomura S, Kurachi O, Furui T, Mizutani S & Tomoda Y. Purification and characterization of 1,25(OH)₂D₃ receptor from human placenta. *Journal of Obstetrics and Gynaecology* 1995 **21** 631–639.
- Henry HL & Norman AW. Vitamin D: metabolism and biological actions. *Annual Review of Nutrition* 1984 **4** 493–520. (doi:10.1146/annurev.nu.04.070184.002425)

- 23 Pérez-Fernandez R, Alonso M, Segura C, Muñoz I, García-Caballero T & Diguez C. Vitamin D receptor gene expression in human pituitary gland. *Life Sciences* 1997 **60** 35–42. (doi:10.1016/S0024-3205(96)00586-3)
- 24 Viganò P, Lattuada D, Mangioni S, Ermellino L, Vignali M, Caporizzo E, Panina-Bordignon P, Besozzi M & Di Blasio AM. Cycling and early pregnant endometrium as a site of regulated expression of the vitamin D system. *Journal of Molecular Endocrinology* 2006 **36** 415–424. (doi:10.1677/jme.1.01946)
- 25 Johnson JA, Grande JP, Roche PC & Kumar R. Immunohistochemical detection and distribution of the 1,25-dihydroxyvitamin D₃ receptor in rat reproductive tissues. *Histochemistry and Cell Biology* 1996 **105** 7–15. (doi:10.1007/BF01450873)
- 26 Merke J, Hügel U & Ritz E. Nuclear testicular 1,25-dihydroxyvitamin D₃ receptors in Sertoli cells and seminiferous tubules of adult rodents. *Biochemical and Biophysical Research Communications* 1985 **127** 303–309. (doi:10.1016/S0006-291X(85)80159-5)
- 27 Habib FK, Maddy SQ & Gelly KJ. Characterisation of receptors for 1,25-dihydroxyvitamin D₃ in the human testis. *Journal of Steroid Biochemistry* 1990 **35** 195–199. (doi:10.1016/0022-4731(90)90274-V)
- 28 Corbett ST, Hill O & Nangia AK. Vitamin D receptor found in human sperm. *Urology* 2006 **68** 1345–1349. (doi:10.1016/j.urology.2006.09.011)
- 29 Aquila S, Guido C, Perrotta I, Tripepi S, Nastro A & Andò S. Human sperm anatomy: ultrastructural localization of 1 α ,25-dihydroxyvitamin D receptor and its possible role in the human male gamete. *Journal of Anatomy* 2008 **213** 555–564. (doi:10.1111/j.1469-7580.2008.00975.x)
- 30 Blomberg Jensen M, Nielsen JE, Jørgensen A, Rajpert-De Meyts E, Kristensen DM, Jørgensen N, Skakkebaek NE, Juul A & Leffers H. Vitamin D receptor and vitamin D metabolizing enzymes are expressed in the human male reproductive tract. *Human Reproduction* 2010 **25** 1303–1311. (doi:10.1093/humrep/deq024)
- 31 Sun T, Zhao Y, Mangelsdorf DJ & Simpson ER. Characterization of a region upstream of exon I.1 of the human CYP19 (aromatase) gene that mediates regulation by retinoids in human choriocarcinoma cells. *Endocrinology* 1998 **139** 1684–1691. (doi:10.1210/en.139.4.1684)
- 32 Barrera D, Avila E, Hernández G, Méndez I, González L, Halhali A, Larrea F, Morales A & Díaz L. Calcitriol affects hCG gene transcription in cultured human syncytiotrophoblasts. *Reproductive Biology and Endocrinology* 2008 **6** 3. (doi:10.1186/1477-7827-6-3)
- 33 Barrera D, Avila E, Hernández G, Halhali A, Biruete B, Larrea F & Díaz L. Estradiol and progesterone synthesis in human placenta is stimulated by calcitriol. *Journal of Steroid Biochemistry and Molecular Biology* 2007 **103** 529–532. (doi:10.1016/j.jsbmb.2006.12.097)
- 34 Tuan RS, Moore CJ, Brittingham JW, Kirwin JJ, Akins RE & Wong M. *In vitro* study of placental trophoblast calcium uptake using JEG-3 human choriocarcinoma cells. *Journal of Cell Science* 1991 **98** 333–342.
- 35 Stephanou A, Ross R & Handwerger S. Regulation of human placental lactogen expression by 1,25-dihydroxyvitamin D₃. *Endocrinology* 1994 **135** 2651–2656. (doi:10.1210/en.135.6.2651)
- 36 Du H, Daftary GS, Lalwani SI & Taylor HS. Direct regulation of HOXA10 by 1,25-(OH)₂D₃ in human myelomonocytic cells and human endometrial stromal cells. *Molecular Endocrinology* 2005 **19** 2222–2233. (doi:10.1210/me.2004-0336)
- 37 Bagot CN, Troy PJ & Taylor HS. Alteration of maternal Hoxa10 expression by *in vivo* gene transfection affects implantation. *Gene Therapy* 2000 **7** 1378–1384. (doi:10.1038/sj.gt.3301245)
- 38 Hirai T, Tsujimura A, Ueda T, Fujita K, Matsuoka Y, Takao T, Miyagawa Y, Koike N & Okuyama A. Effect of 1,25-dihydroxyvitamin D on testicular morphology and gene expression in experimental cryptorchid mouse: testis specific cDNA microarray analysis and potential implication in male infertility. *Journal of Urology* 2009 **181** 1487–1492. (doi:10.1016/j.juro.2008.11.007)
- 39 Selva DM, Hirsch-Reinshagen V, Burgess B, Zhou S, Chan J, McIsaac S, Hayden MR, Hammond GL, Vogl AW & Wellington CL. The ATP-binding cassette transporter 1 mediates lipid efflux from Sertoli cells and influences male fertility. *Journal of Lipid Research* 2004 **45** 1040–1050. (doi:10.1194/jlr.M400007-JLR200)
- 40 Zanatta L, Zamoner A, Gonçalves R, Zanatta AP, Bouraïma-Lelong H, Bois C, Carreau S & Silva FR. Effect of 1 α ,25-dihydroxyvitamin D(3) in plasma membrane targets in immature rat testis: ionic channels and γ -glutamyl transpeptidase activity. *Archives of Biochemistry and Biophysics* 2011 **515** 46–53. (doi:10.1016/j.abb.2011.09.001)
- 41 Zanatta L, Zamoner A, Zanatta AP, Bouraïma-Lelong H, Delalande C, Bois C, Carreau S & Silva FR. Nongenomic and genomic effects of 1 α ,25(OH)₂ vitamin D₃ in rat testis. *Life Sciences* 2011 **89** 515–523. (doi:10.1016/j.lfs.2011.04.008)
- 42 Nangia V, Matin A, Bhojwani K, Kulkarni M, Yadav M & Jonas JB. Testicular maturation arrest to testis cancer: spectrum of expression of the vitamin D receptor and vitamin D treatment *in vitro*. *Journal of Urology* 2007 **178** 1092–1096. (doi:10.1016/j.juro.2007.05.009)
- 43 Somjen D, Katzburg S, Stern N, Kohen F, Sharon O, Limor R, Jaccard N, Hendel D & Weisman Y. 25 Hydroxy-vitamin D(3)-1 α hydroxylase expression and activity in cultured human osteoblasts and their modulation by parathyroid hormone, estrogenic compounds and dihydrotestosterone. *Journal of Steroid Biochemistry and Molecular Biology* 2007 **107** 238–244. (doi:10.1016/j.jsbmb.2007.03.048)
- 44 Menegaz D, Rosso A, Royer C, Leite LD, Santos AR & Silva FR. Role of 1 α ,25(OH)₂ vitamin D₃ on α -[1-(14)C]MeAIB accumulation in immature rat testis. *Steroids* 2009 **74** 264–269. (doi:10.1016/j.steroids.2008.11.015)
- 45 Akerstrom VL & Walters MR. Physiological effects of 1,25-dihydroxyvitamin D₃ in TM4 Sertoli cell line. *American Journal of Physiology* 1992 **262** E884–E889.
- 46 Aquila S, Guido C, Middea E, Perrotta I, Bruno R, Pellegrino M & Andò S. Human male gamete endocrinology: 1 α , 25-dihydroxyvitamin D₃ (1,25(OH)₂D₃) regulates different aspects of human sperm biology and metabolism. *Reproductive Biology and Endocrinology* 2009 **7** 140. (doi:10.1186/1477-7827-7-140)
- 47 De Felici M, Dolci S & Siracusa G. An increase of intracellular free Ca²⁺ is essential for spontaneous meiotic resumption by mouse oocytes. *Journal of Experimental Zoology* 1991 **260** 401–405. (doi:10.1002/jez.1402600314)
- 48 Halhali A, Acker GM & Garabédian M. 1,25-Dihydroxyvitamin D₃ induces *in vivo* the decidualization of rat endometrial cells. *Journal of Reproduction and Fertility* 1991 **91** 59–64. (doi:10.1530/jrf.0.0910059)
- 49 Horii I, Takizawa S & Fujii T. Effect of 1,25-dihydroxyvitamin D₃ on the female reproductive system in rats. *Journal of Toxicological Sciences* 1992 **17** 91–105. (doi:10.2131/jts.17.91)
- 50 Hamden K, Carreau S, Jamoussi K, Ayadi F, Garmazi F, Mezgeni N & Elfeki A. Inhibitory effects of 1 α , 25dihydroxyvitamin D₃ and Ajuga iva extract on oxidative stress, toxicity and hypo-fertility in diabetic rat testes. *Journal of Physiology and Biochemistry* 2008 **64** 231–239. (doi:10.1007/BF03216108)
- 51 Kwiecinski GG, Petrie GI & DeLuca HF. 1,25-Dihydroxyvitamin D₃ restores fertility of vitamin D-deficient female rats. *American Journal of Physiology* 1989 **256** E483–E487.
- 52 Kwiecinski GG, Petrie GI & DeLuca HF. Vitamin D is necessary for reproductive functions of the male rat. *Journal of Nutrition* 1989 **119** 741–744.
- 53 Osmundsen BC, Huang HF, Anderson MB, Christakos S & Walters MR. Multiple sites of action of the vitamin D endocrine system: FSH stimulation of testis 1,25-dihydroxyvitamin D₃ receptors. *Journal of Steroid Biochemistry* 1989 **34** 339–343. (doi:10.1016/0022-4731(89)90105-2)

- 54 Umland AM, Kwiecinski GG & DeLuca HF. Normalization of serum calcium restores fertility in vitamin D-deficient male rats. *Journal of Nutrition* 1992 **122** 1338–1344.
- 55 Yoshizawa T, Handa Y, Uematsu Y, Takeda S, Sekine K, Yoshihara Y, Kawakami T, Arioka K, Sato H, Uchiyama Y, Masushige S, Fukamizu A, Matsumoto T & Kato S. Mice lacking the vitamin D receptor exhibit impaired bone formation, uterine hypoplasia and growth retardation after weaning. *Nature Genetics* 1997 **16** 391–396. (doi:10.1038/ng0897-391)
- 56 Kovacs CS, Woodland ML, Fudge NJ & Friel JK. The vitamin D receptor is not required for fetal mineral homeostasis or for the regulation of placental calcium transfer in mice. *American Journal of Physiology, Endocrinology and Metabolism* 2005 **289** E133–E144. (doi:10.1152/ajpendo.00354.2004)
- 57 Panda DK, Miao D, Tremblay ML, Sirois J, Farookhi R, Hendy GN & Goltzman D. Targeted ablation of the 25-hydroxyvitamin D 1 α -hydroxylase enzyme: evidence for skeletal, reproductive, and immune dysfunction. *PNAS* 2001 **98** 7498–7503. (doi:10.1073/pnas.131029498)
- 58 Johnson LE & DeLuca HF. Vitamin D receptor null mutant mice fed high levels of calcium are fertile. *Journal of Nutrition* 2001 **131** 1787–1791.
- 59 Sun W, Xie H, Ji J, Zhou X, Goltzman D & Miao D. Defective female reproductive function in 1,25(OH)₂D-deficient mice results from indirect effect mediated by extracellular calcium and/or phosphorus. *American Journal of Physiology, Endocrinology and Metabolism* 2010 **299** E928–E935. (doi:10.1152/ajpendo.00378.2010)
- 60 Rojansky N, Brzezinski A & Schenker JG. Seasonality in human reproduction: an update. *Human Reproduction* 1992 **7** 735–745.
- 61 Rojansky N, Benshushan A, Meirsdorf S, Lewin A, Laufer N & Safran A. Seasonal variability in fertilization and embryo quality rates in women undergoing IVF. *Fertility and Sterility* 2000 **74** 476–481. (doi:10.1016/S0015-0282(00)00669-5)
- 62 Ozkan S, Jindal S, Greenseed K, Shu J, Zeitlian G, Hickmon C & Pal L. Replete vitamin D stores predict reproductive success following *in vitro* fertilization. *Fertility and Sterility* 2009 **94** 1314–1319. (doi:10.1016/j.fertnstert.2009.05.019)
- 63 Aleyasin A, Hosseini MA, Mahdavi A, Safdarian L, Fallahi P, Mohajeri MR, Abbasi M & Esfahani F. Predictive value of the level of vitamin D in follicular fluid on the outcome of assisted reproductive technology. *European Journal of Obstetrics, Gynecology, and Reproductive Biology* 2011 **159** 132–137. (doi:10.1016/j.ejogrb.2011.07.006)
- 64 Anifandis GM, Dafopoulos K, Messini CI, Chalvatzas N, Liakos N, Pournaras S & Messinis IE. Prognostic value of follicular fluid 25-OH vitamin D and glucose levels in the IVF outcome. *Reproductive Biology and Endocrinology* 2010 **8** 91. (doi:10.1186/1477-7827-8-91)
- 65 Asunción M, Calvo RM, San Millán JL, Sancho J, Avila S & Escobar-Morreale HF. A prospective study of the prevalence of the polycystic ovary syndrome in unselected Caucasian women from Spain. *Journal of Clinical Endocrinology and Metabolism* 2000 **85** 2434–2438. (doi:10.1210/jc.85.7.2434)
- 66 Diamanti-Kandarakis E, Kouli CR, Bergiele AT, Filandra FA, Tsianateli TC, Spina GG, Zapanti ED & Bartzis MI. A survey of the polycystic ovary syndrome in the Greek island of Lesbos: hormonal and metabolic profile. *Journal of Clinical Endocrinology and Metabolism* 1999 **84** 4006–4011. (doi:10.1210/jc.84.11.4006)
- 67 Dunaif A. Insulin resistance and the polycystic ovary syndrome: mechanism and implications for pathogenesis. *Endocrine Reviews* 1997 **18** 774–800. (doi:10.1210/er.18.6.774)
- 68 Wehr E, Möller R, Horejsi R, Giuliani A, Kopera D, Schweighofer N, Groselj-Strele A, Pieber TR & Obermayer-Pietsch B. Subcutaneous adipose tissue topography and metabolic disturbances in polycystic ovary syndrome. *Wiener Klinische Wochenschrift* 2009 **121** 262–269. (doi:10.1007/s00508-009-1162-2)
- 69 Ehrmann DA. Polycystic ovary syndrome. *New England Journal of Medicine* 2005 **352** 1223–1236. (doi:10.1056/NEJMra041536)
- 70 Wehr E, Pilz S, Schweighofer N, Giuliani A, Kopera D, Pieber TR & Obermayer-Pietsch B. Association of hypovitaminosis D with metabolic disturbances in polycystic ovary syndrome. *European Journal of Endocrinology* 2009 **161** 575–582. (doi:10.1530/EJE-09-0432)
- 71 Ngo DT, Chan WP, Rajendran S, Heresztyn T, Amarasekera A, Sverdlow AL, O'Loughlin PD, Morris HA, Chirkov YY, Norman RJ & Horowitz JD. Determinants of insulin responsiveness in young women: impact of polycystic ovarian syndrome, nitric oxide, and vitamin D. *Nitric Oxide* 2011 **25** 326–330. (doi:10.1016/j.niox.2011.06.005)
- 72 Yildizhan R, Kurdoglu M, Adali E, Kulusari A, Yildizhan B, Sahin HG & Kamaci M. Serum 25-hydroxyvitamin D concentrations in obese and non-obese women with polycystic ovary syndrome. *Archives of Gynecology and Obstetrics* 2009 **280** 559–563. (doi:10.1007/s00404-009-0958-7)
- 73 Hahn S, Haselhorst U, Tan S, Quadbeck B, Schmidt M, Roesler S, Kimmig R, Mann K & Janssen OE. Low serum 25-hydroxyvitamin D concentrations are associated with insulin resistance and obesity in women with polycystic ovary syndrome. *Experimental and Clinical Endocrinology and Diabetes* 2006 **114** 577–583. (doi:10.1055/s-2006-948308)
- 74 Mahmoudi T, Gourabi H, Ashrafi M, Yazdi RS & Ezabadi Z. Calcitropic hormones, insulin resistance, and the polycystic ovary syndrome. *Fertility and Sterility* 2010 **93** 1208–1214. (doi:10.1016/j.fertnstert.2008.11.031)
- 75 Li HW, Brereton RE, Anderson RA, Wallace AM & Ho CK. Vitamin D deficiency is common and associated with metabolic risk factors in patients with polycystic ovary syndrome. *Metabolism* 2011 **60** 1475–1481. (doi:10.1016/j.metabol.2011.03.002)
- 76 Panidis D, Balaris C, Farmakiotis D, Rousso D, Kourtis A, Balaris V, Katsikis I, Zournatzi V & Diamanti-Kandarakis E. Serum parathyroid hormone concentrations are increased in women with polycystic ovary syndrome. *Clinical Chemistry* 2005 **51** 1691–1697. (doi:10.1373/clinchem.2005.052761)
- 77 Chavarro JE, Rich-Edwards JW, Rosner B & Willett WC. A prospective study of dairy foods intake and anovulatory infertility. *Human Reproduction* 2007 **22** 1340–1347. (doi:10.1093/humrep/dem019)
- 78 Wortsman J, Matsuoka LY, Chen TC, Lu Z & Holick MF. Decreased bioavailability of vitamin D in obesity. *American Journal of Clinical Nutrition* 2000 **72** 690–693.
- 79 Compston JE, Vedi S, Ledger JE, Webb A, Gazet JC & Pilkington TR. Vitamin D status and bone histomorphometry in gross obesity. *American Journal of Clinical Nutrition* 1981 **34** 2359–2363.
- 80 Kamycheva E, Joakimsen RM & Jorde R. Intakes of calcium and vitamin D predict body mass index in the population of Northern Norway. *Journal of Nutrition* 2003 **133** 102–106.
- 81 Pittas AG, Lau J, Hu FB & Dawson-Hughes B. The role of vitamin D and calcium in type 2 diabetes. A systematic review and meta-analysis. *Journal of Clinical Endocrinology and Metabolism* 2007 **92** 2017–2029. (doi:10.1210/jc.2007-0298)
- 82 Maestro B, Dávila N, Carranza MC & Calle C. Identification of a vitamin D response element in the human insulin receptor gene promoter. *Journal of Steroid Biochemistry and Molecular Biology* 2003 **84** 223–230. (doi:10.1016/S0960-0760(03)00032-3)
- 83 Maestro B, Molero S, Bajo S, Dávila N & Calle C. Transcriptional activation of the human insulin receptor gene by 1,25-dihydroxyvitamin D(3). *Cell Biochemistry and Function* 2002 **20** 227–232. (doi:10.1002/cbf.951)
- 84 Milner RD & Hales CN. The role of calcium and magnesium in insulin secretion from rabbit pancreas studied *in vitro*. *Diabetologia* 1967 **3** 47–49. (doi:10.1007/BF01269910)
- 85 Bikle D. Nonclassic actions of vitamin D. *Journal of Clinical Endocrinology and Metabolism* 2009 **94** 26–34. (doi:10.1210/jc.2008-1454)
- 86 Shoelson SE, Herrero L & Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology* 2007 **132** 2169–2180. (doi:10.1053/j.gastro.2007.03.059)

- 87 Banaszewska B, Pawelczyk L, Spaczynski RZ & Duleba AJ. Effects of simvastatin and metformin on polycystic ovary syndrome after six months of treatment. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** 3494–3501. (doi:10.1210/jc.2011-0501)
- 88 Sathyapalan T, Shepherd J, Arnett C, Coady AM, Kilpatrick ES & Atkin SL. Atorvastatin increases 25-hydroxy vitamin D concentrations in patients with polycystic ovary syndrome. *Clinical Chemistry* 2010 **56** 1696–1700. (doi:10.1373/clinchem.2010.144014)
- 89 Mahmoudi T. Genetic variation in the vitamin D receptor and polycystic ovary syndrome risk. *Fertility and Sterility* 2009 **92** 1381–1383. (doi:10.1016/j.fertnstert.2009.05.002)
- 90 Wehr E, Trummer O, Giuliani A, Gruber HJ, Pieber TR & Obermayer-Pietsch B. Vitamin D-associated polymorphisms are related to insulin resistance and vitamin D deficiency in polycystic ovary syndrome. *European Journal of Endocrinology* 2011 **164** 741–749. (doi:10.1530/EJE-11-0134)
- 91 Ranjzad F, Mahban A, Shemirani AI, Mahmoudi T, Vahedi M, Nikzamir A & Zali MR. Influence of gene variants related to calcium homeostasis on biochemical parameters of women with polycystic ovary syndrome. *Journal of Assisted Reproduction and Genetics* 2011 **28** 225–232. (doi:10.1007/s10815-010-9506-4)
- 92 Wang TJ, Zhang F, Richards JB, Kestenbaum B, van Meurs JB, Berry D, Kiel DP, Streeten EA, Ohlsson C, Koller DL, Peltonen L, Cooper JD, O'Reilly PF, Houston DK, Glazer NL, Vandenput L, Peacock M, Shi J, Rivadeneira F, McCarthy MI, Anneli P, de Boer IH, Mangino M, Kato B, Smyth DJ, Booth SL, Jacques PF, Burke GL, Goodarzi M, Cheung CL, Wolf M, Rice K, Goltzman D, Hidioglou N, Ladouceur M, Wareham NJ, Hocking LJ, Hart D, Arden NK, Cooper C, Malik S, Fraser WD, Hartikainen AL, Zhai G, Macdonald HM, Forouhi NG, Loos RJ, Reid DM, Hakim A, Dennison E, Liu Y, Power C, Stevens HE, Jaana L, Vasani RS, Soranzo N, Bojunga J, Psaty BM, Lorentzon M, Forouhi T, Harris TB, Hofman A, Jansson JO, Cauley JA, Uitterlinden AG, Gibson Q, Järvelin MR, Karasik D, Siscovick DS, Econs MJ, Kritchevsky SB, Florez JC, Todd JA, Dupuis J, Hyppönen E & Spector TD. Common genetic determinants of vitamin D insufficiency: a genome-wide association study. *Lancet* 2010 **376** 180–188. (doi:10.1016/S0140-6736(10)60588-0)
- 93 Thys-Jacobs S, Donovan D, Papadopoulos A, Sarrel P & Bilezikian JP. Vitamin D and calcium dysregulation in the polycystic ovarian syndrome. *Steroids* 1999 **64** 430–435. (doi:10.1016/S0039-128X(99)00012-4)
- 94 Selimoglu H, Duran C, Kiyici S, Ersoy C, Guclu M, Ozkaya G, Tuncel E, Erturk E & Imamoglu S. The effect of vitamin D replacement therapy on insulin resistance and androgen levels in women with polycystic ovary syndrome. *Journal of Endocrinological Investigation* 2010 **33** 234–238.
- 95 Kotsa K, Yavropoulou MP, Anastasiou O & Yovos JG. Role of vitamin D treatment in glucose metabolism in polycystic ovary syndrome. *Fertility and Sterility* 2009 **92** 1053–1058. (doi:10.1016/j.fertnstert.2008.07.1757)
- 96 Wehr E, Pieber TR & Obermayer-Pietsch B. Effect of vitamin D₃ treatment on glucose metabolism and menstrual frequency in PCOS women – a pilot study. *Journal of Endocrinological Investigation* 2011 **34** 757–763. (doi:10.3275/7748)
- 97 Rashidi B, Haghollahi F, Shariat M & Zayerii F. The effects of calcium-vitamin D and metformin on polycystic ovary syndrome: a pilot study. *Taiwanese Journal of Obstetrics and Gynecology* 2009 **48** 142–147. (doi:10.1016/S1028-4559(09)60275-8)
- 98 Knight JA, Wong J, Blackmore KM, Raboud JM & Vieth R. Vitamin D association with estradiol and progesterone in young women. *Cancer Causes and Control* 2010 **21** 479–483. (doi:10.1007/s10552-009-9466-0)
- 99 Freedman DM, Looker AC, Chang SC & Graubard BI. Prospective study of serum vitamin D and cancer mortality in the United States. *Journal of the National Cancer Institute* 2007 **99** 1594–1602. (doi:10.1093/jnci/djm204)
- 100 Somigliana E, Panina-Bordignon P, Murone S, Di Lucia P, Vercellini P & Vigano P. Vitamin D reserve is higher in women with endometriosis. *Human Reproduction* 2007 **22** 2273–2278. (doi:10.1093/humrep/dem142)
- 101 Hartwell D, Rødbro P, Jensen SB, Thomsen K & Christiansen C. Vitamin D metabolites – relation to age, menopause and endometriosis. *Scandinavian Journal of Clinical and Laboratory Investigation* 1990 **50** 115–121. (doi:10.3109/00365519009089142)
- 102 Vilarino FL, Bianco B, Lerner TG, Teles JS, Mafra FA, Christofolini DM & Barbosa CP. Analysis of vitamin D receptor gene polymorphisms in women with and without endometriosis. *Human Immunology* 2011 **72** 359–363. (doi:10.1016/j.humimm.2011.01.006)
- 103 Faserl K, Golderer G, Kremser L, Lindner H, Sarg B, Wildt L & Seeber B. Polymorphism in vitamin D-binding protein as a genetic risk factor in the pathogenesis of endometriosis. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** E233–E241. (doi:10.1210/jc.2010-1532)
- 104 Borkowski J, Gmyrek GB, Madej JP, Nowacki W, Goluda M, Gabryś M, Stefaniak T & Chelmońska-Soyta A. Serum and peritoneal evaluation of vitamin D-binding protein in women with endometriosis. *Postepy Higieny i Medycyny Doswiadczalnej* 2008 **62** 103–111.
- 105 Ferrero S, Gillott DJ, Anserini P, Remorgida V, Price KM, Ragni N & Grudzinskas JG. Vitamin D binding protein in endometriosis. *Journal of the Society of Gynecological Investigation* 2005 **12** 272–277. (doi:10.1016/j.jsjg.2005.01.027)
- 106 Yoshida M, Kawano N & Yoshida K. Control of sperm motility and fertility: diverse factors and common mechanisms. *Cellular and Molecular Life Sciences* 2008 **65** 3446–3457. (doi:10.1007/s00018-008-8230-z)
- 107 Blomberg Jensen M, Bjerrum PJ, Jessen TE, Nielsen JE, Joensen UN, Olesen IA, Petersen JH, Juul A, Dissing S & Jørgensen N. Vitamin D is positively associated with sperm motility and increases intracellular calcium in human spermatozoa. *Human Reproduction* 2011 **26** 1307–1317. (doi:10.1093/humrep/der059)
- 108 Ramlau-Hansen CH, Moeller UK, Bonde JP, Olsen J & Thulstrup AM. Are serum levels of vitamin D associated with semen quality? Results from a cross-sectional study in young healthy men. *Fertility and Sterility* 2011 **95** 1000–1004. (doi:10.1016/j.fertnstert.2010.11.002)
- 109 Foresta C, Strapazzon G, De Toni L, Perilli L, Di Mambro A, Muciaccia B, Sartori L & Selice R. Bone mineral density and testicular failure: evidence for a role of vitamin D 25-hydroxylase in human testis. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** E646–E652. (doi:10.1210/jc.2010-1628)
- 110 Tiwari A. Elocalcitol, a vitamin D₃ analog for the potential treatment of benign prostatic hyperplasia, overactive bladder and male infertility. *IDrugs: the Investigational Drugs Journal* 2009 **12** 2040–2410.
- 111 Wehr E, Pilz S, Boehm BO, März W, Grammer TB & Obermayer-Pietsch B. Sex steroids and mortality in men referred for coronary angiography. *Clinical Endocrinology* 2010 **73** 613–621. (doi:10.1111/j.1365-2265.2010.03852.x)
- 112 Wehr E, Pilz S, Boehm BO, März W, Grammer T & Obermayer-Pietsch B. Low free testosterone is associated with heart failure mortality in older men referred for coronary angiography. *European Journal of Heart Failure* 2011 **13** 482–488. (doi:10.1093/eurjhf/hfr007)
- 113 Glass AR, Swerdloff RS, Bray GA, Dahms WT & Atkinson RL. Low serum testosterone and sex-hormone-binding-globulin in massively obese men. *Journal of Clinical Endocrinology and Metabolism* 1977 **45** 1211–1219. (doi:10.1210/jcem-45-6-1211)
- 114 Mordan-McCombs S, Brown T, Wang WL, Gaupel AC, Welsh J & Tenniswood M. Tumor progression in the LPB-Tag transgenic model of prostate cancer is altered by vitamin D receptor and serum testosterone status. *Journal of Steroid Biochemistry and Molecular Biology* 2010 **121** 368–371. (doi:10.1016/j.jsbmb.2010.03.062)

- 115 Lee DM, Tajar A, Pye SR, Boonen S, Vanderschueren D, Bouillon R, O'Neill TW, Bartfai G, Casanueva FF, Finn JD, Forti G, Giwercman A, Han TS, Huhtaniemi IT, Kula K, Lean ME, Pendleton N, Punab M & Wu FC & the EMAS study group. Association of hypogonadism with vitamin D status: the European Male Ageing Study. *European Journal of Endocrinology* 2012 **166** 77–85. (doi:10.1530/EJE-11-0743)
- 116 Dent CE & Gupta MM. Plasma 25-hydroxyvitamin-D-levels during pregnancy in Caucasians and in vegetarian and non-vegetarian Asians. *Lancet* 1975 **2** 1057–1060. (doi:10.1016/S0140-6736(75)90430-4)
- 117 Turtton CW, Stanley P, Stamp TC & Maxwell JD. Altered vitamin-D metabolism in pregnancy. *Lancet* 1977 **1** 222–225. (doi:10.1016/S0140-6736(77)91017-0)
- 118 Looker AC, Pfeiffer CM, Lacher DA, Schleicher RL, Picciano MF & Yetley EA. Serum 25-hydroxyvitamin D status of the US population: 1988–1994 compared with 2000–2004. *American Journal of Clinical Nutrition* 2008 **88** 1519–1527. (doi:10.3945/ajcn.2008.26182)
- 119 Zhang C, Qiu C, Hu FB, David RM, van Dam RM, Bralley A & Williams MA. Maternal plasma 25-hydroxyvitamin D concentrations and the risk for gestational diabetes mellitus. *PLoS ONE* 2008 **3** e3753. (doi:10.1371/journal.pone.0003753)
- 120 Maghbooli Z, Hossein-Nezhad A, Karimi F, Shafaei AR & Larijani B. Correlation between vitamin D₃ deficiency and insulin resistance in pregnancy. *Diabetes Metabolism Research and Reviews* 2008 **24** 27–32. (doi:10.1002/dmrr.737)
- 121 Bodnar LM, Catov JM, Simhan HN, Holick MF, Powers RW & Roberts JM. Maternal vitamin D deficiency increases the risk of preeclampsia. *Journal of Clinical Endocrinology and Metabolism* 2007 **92** 3517–3522. (doi:10.1210/jc.2007-0718)
- 122 Bodnar LM, Krohn MA & Simhan HN. Maternal vitamin D deficiency is associated with bacterial vaginosis in the first trimester of pregnancy. *Journal of Nutrition* 2009 **139** 1157–1161. (doi:10.3945/jn.108.103168)
- 123 Haugen M, Brantsaeter AL, Trogstad L, Alexander J, Roth C, Magnus P & Meltzer HM. Vitamin D supplementation and reduced risk of preeclampsia in nulliparous women. *Epidemiology* 2009 **20** 720–726. (doi:10.1097/EDE.0b013e3181a70f08)
- 124 Bodnar LM, Catov JM, Zmuda JM, Cooper ME, Parrott MS, Roberts JM, Marazita ML & Simhan HN. Maternal serum 25-hydroxyvitamin D concentrations are associated with small-for-gestational age births in white women. *Journal of Nutrition* 2010 **140** 999–1006. (doi:10.3945/jn.109.119636)
- 125 Brooke OG, Brown IR, Bone CD, Carter ND, Cleeve HJ, Maxwell JD, Robinson VP & Winder SM. Vitamin D supplements in pregnant Asian women: effects on calcium status and fetal growth. *BMJ* 1980 **280** 751–754. (doi:10.1136/bmj.280.6216.751)
- 126 Wagner CL & Greer FR & American Academy of Pediatrics Section on Breastfeeding; American Academy of Pediatrics Committee on Nutrition. Prevention of rickets and vitamin D deficiency in infants, children, and adolescents. *Pediatrics* 2008 **122** 1142–1152. (doi:10.1542/peds.2008-1862)
- 127 Javaid MK, Crozier SR, Harvey NC, Gale CR, Dennison EM, Boucher BJ, Arden NK, Godfrey KM & Cooper C & Princess Anne Hospital Study Group. Maternal vitamin D status during pregnancy and childhood bone mass at age 9 years: a longitudinal study. *Lancet* 2006 **367** 36–43. (doi:10.1016/S0140-6736(06)67922-1)
- 128 Camargo CA Jr, Rifas-Shiman SL, Litonjua AA, Rich-Edwards JW, Weiss ST, Gold DR, Kleinman K & Gillman MW. Maternal intake of vitamin D during pregnancy and risk of recurrent wheeze in children at 3 y of age. *American Journal of Clinical Nutrition* 2007 **85** 788–795.
- 129 McGrath J. Does 'imprinting' with low prenatal vitamin D contribute to the risk of various adult disorders? *Medical Hypotheses* 2001 **56** 367–371. (doi:10.1054/mehy.2000.1226)
- 130 Hollis BW, Johnson D, Hulsey TC, Ebeling M & Wagner CL. Vitamin D supplementation during pregnancy: double-blind, randomized clinical trial of safety and effectiveness. *Journal of Bone and Mineral Research* 2011 **26** 2341–2357. (doi:10.1002/jbmr.463)
- 131 Hollis BW. Randomized controlled trials to determine the safety of vitamin D supplementation during pregnancy and lactation. In *Fourteenth Workshop on Vitamin D*. Brugge, Belgium, 2009.
- 132 Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH & Weaver CM. Evaluation, treatment, and prevention of vitamin D deficiency: an endocrine society clinical practice guideline. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** 1911–1930. (doi:10.1210/jc.2011-0385)
- 133 Heany RP. Vitamin D in health and disease. *Clinical Journal of the American Society of Nephrology* 2008 **3** 1535–1541. (doi:10.2215/CJN.01160308)
- 134 Chapuy MC, Preziosi P, Maamer M, Arnaud S, Galan P, Hercberg S & Meunier PJ. Prevalence of vitamin D insufficiency in an adult normal population. *Osteoporosis International* 1997 **7** 439–443. (doi:10.1007/s001980050030)

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